

# The Effects of Rising Temperature on the Ecophysiology of Tropical Forest Trees

Martijn Slot and Klaus Winter

**Abstract** The response of tropical trees to rising temperatures represents a key uncertainty that limits our ability to predict biosphere-atmosphere feedbacks in a warming world. We review the current understanding of temperature effects on the ecophysiology of tropical trees from organelle to biome level, where we distinguish between short-term responses, acclimation, and adaptation. We present new data on short-term temperature responses of photosynthesis and dark respiration, and temperature acclimation of photosynthesis. We also compare new field and laboratory-obtained photosynthesis-temperature response data. We identify several priority study areas. (1) Acclimation: We need to better understand photosynthetic acclimation, for example to determine whether the adjustment of the thermal optimum of photosynthesis ( $T_{Opt}$ ) is consistently negated by a decrease in photosynthesis at  $T_{Opt}$ , as we observed. (2) Growth: Whereas tropical seedlings may grow better with warming, canopy trees reportedly grow worse; we do not currently know what explains these contrasting temperature effects. (3) Reproduction: Tropical trees may be close to reproductive temperature thresholds, as heat sterility in crops occurs in the upper 30 °C range. Nonetheless, the temperature sensitivity of tropical tree reproduction is virtually unstudied. (4) Mortality: How does heat-induced atmospheric drought (high leaf-to-air vapor pressure deficit) affect tropical tree mortality? (5) Stomatal behavior: What is the specific role of temperature in the induction of midday-stomatal closure on sunny days? Better knowledge in these areas will improve our ability to predict carbon fluxes in tropical forests experiencing ongoing warming.

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M. Slot (✉) · K. Winter  
Smithsonian Tropical Research Institute, Apartado 0843-03092,  
Balboa, Ancón, Republic of Panama  
e-mail: SlotM@si.edu; martijnslot78@gmail.com

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## Introduction

Tropical forests cover only 15 % of the planet's terrestrial surface (Pan et al. 2013), yet they account for more than one-third of its net primary productivity (NPP) (Saugier et al. 2001) and two-thirds of its plant biomass (Pan et al. 2013). Given this disproportionately large contribution to the global carbon cycle, it is important that we increase our understanding of the effects of global warming on tropical forest trees. Recent analyses have suggested that global variation in temperature and precipitation have no direct effect on global patterns of NPP after accounting for stand age and biomass (Michaletz et al. 2014), but this does not mean that changes in climatic variables will not affect NPP within a given biome. The coming decades will see ongoing warming in tropical regions, leading to unprecedented temperature regimes (Diffenbaugh and Scherer 2011) that currently do not support closed-canopy forests (Wright et al. 2009). Tropical regions have previously experienced warming—most notably leading up to the Paleocene-Eocene Thermal Maximum (~56 million years ago) when temperatures rose by 3–5 °C—but even the most rapid of such historical warming events took place over thousands to tens of thousands of years, timescales during which gradual changes in species composition, adaptive responses, and speciation are possible (Jaramillo et al. 2010). Current warming, on the other hand, occurs over the lifetime of individual trees, necessitating a high degree of thermal plasticity to maintain long-term growth and survival.

Warming effects on the physiology of woody plants have been studied primarily in mid- and high-latitude ecosystems. Many of these effects are expected to be broadly generalizable, given the universal effects of temperature on enzyme kinetics, and the common principles of the biochemical pathways of both photosynthesis and mitochondrial respiration, the key drivers of terrestrial biosphere-atmosphere carbon exchange. However, there are several important differences between tropical and higher latitude ecosystems that might differentially affect plant responses to climate warming.

First, seasonal temperature fluctuations are minimal in the tropics, and such thermally stable conditions may not favor evolution of the capacity to acclimate to temperature changes (Janzen 1967; Cunningham and Read 2003). If tropical species have limited thermal plasticity, current warming may be highly detrimental to their performance. Second, for most of the past 2.6 million years tropical regions have experienced lower temperatures than today. Natural selection under such conditions would not have favored heat-protective traits (Corlett 2011). Temperate and boreal climates have also been cooler over this period, but current warming is more likely to expose tropical vegetation to temperatures near the thermal limit of photosynthesis than higher latitude vegetation. Indeed, tropical forests might be close to their thermal optimum already (Doughty and Goulden 2008; but see Lloyd and Farquhar 2008). Third, warming does not lengthen the growing season in the tropics as it does at higher latitudes (Menzel and Fabian 1999; Menzel et al. 2006).

This means that a longer growing period will not compensate for any negative effect of temperature on carbon uptake in the tropics. Given the above three points, temperature relationships of tropical trees deserve special attention.

Here we review the current understanding of the effects of elevated temperature on the ecophysiology of tropical trees and forests. We present new data on instantaneous effects and growth-temperature effects on seedling gas exchange; we compare laboratory-based measurements with in situ measurements of temperature effects on foliar physiology; we review experimental, observational, and modeling studies; and we identify areas of study that should be prioritized to improve our ability to predict carbon fluxes in gradually warming tropical forests. We will focus on temperature responses at the ecophysiological level, emphasizing the effects of warming in the non-damaging temperature range. For information on molecular responses to heat stress we refer readers to several reviews on this subject that have been published in the past decade (e.g., Wahid et al. 2007; Allakhverdiev et al. 2008; Ashraf and Harris 2013). Our primary concern here is the effect that warming has on tropical lowlands, which are currently the warmest ecosystems that support closed-canopy forests (Wright et al. 2009). The ~15 % of tropical forests that are montane are beyond the scope of this chapter.

We distinguish between different temporal scales at which temperature may affect tropical trees, and between the organizational scales at which these temperature response processes operate (Table 1). We will start our review at the organelle and leaf level—with a strong focus on photosynthesis and respiration—then work our way up through the whole-tree level out to stand-, ecosystem-, and biome-level temperature effects. Along the way we go from a highly mechanistic understanding of organelle- and leaf-level processes—based on foundational work on temperate species and backed up by a large body of experimental work—to more speculation as we attempt to predict temperature effects at higher organizational scales and over longer time periods for which no experimental work exists to date.

## Leaf-Level Temperature Effects

### *Photosynthesis*

Net photosynthesis ( $A_{\text{Net}}$ ) increases with short-term warming before reaching a maximum  $\text{CO}_2$  assimilation rate ( $A_{\text{Opt}}$ ) at optimum temperature ( $T_{\text{Opt}}$ ), beyond which net  $\text{CO}_2$  uptake rates decline and eventually drop to zero at the upper  $\text{CO}_2$  compensation point (Box 1). We will first discuss the short-term temperature response over non-harmful, reversible temperature ranges. After briefly addressing heat damage, we discuss temperature responses at timescales over which acclimation may occur.

**Table 1** Temperature responses (+ and – indicate the direction of change) across organizational and temporal scales

Organizational scale	Biophysical response	Acclimation	Adaptation	Adaptation/Species turnover	Species turnover/speciation
<i>Biome</i>					Possible temperature/drought-induced collapse of closed-canopy forest If closed canopy forest is lost: big changes in rainfall patterns, climate feedbacks; risk of runaway warming
<i>Ecosystem</i>		<ul style="list-style-type: none"> <li>+ Mineralization in soils could benefit tree growth in nitrogen limited ecosystems</li> <li>– NPP/GPP ratio<sup>a</sup></li> </ul>	<ul style="list-style-type: none"> <li>+ Isoprene emission<sup>h</sup></li> <li>– NPP/GPP ratio<sup>a</sup></li> </ul>		Savanna or desert species may take over If closed canopy forest is lost: big changes in hydraulics, rainfall patterns, climate feedbacks; risk of runaway warming
<i>Tree</i>	<ul style="list-style-type: none"> <li>+ Respiration of all organs (however: root and bulk stem temperatures are well buffered and do not track instantaneous air temperature changes)</li> <li>– Pollen viability</li> </ul>	<ul style="list-style-type: none"> <li>– Respiratory capacity of all organs; species-specific interactions with mycorrhizae may either strengthen or weaken the acclimation response<sup>b</sup></li> <li>– Reproductive efficiency</li> <li>– Stem diameter growth of canopy trees</li> <li>Seedling/sapling growth unaffected</li> </ul>	<ul style="list-style-type: none"> <li>Possible change in biomass allocation and leaf display</li> <li>+ Leaf cooling traits for adapted species</li> <li>+ Isoprene emission<sup>h</sup></li> </ul>		If closed canopy forest is lost: increased abundance of savanna or desert species that are better adapted to drought
<i>Leaf</i>	<ul style="list-style-type: none"> <li>Photosynthesis peaks near local mean temperature and then declines</li> <li>+ Respiration</li> <li>+ Isoprene emission</li> <li>+ Monoterpene emission</li> </ul>	<ul style="list-style-type: none"> <li>+ <math>T_{Opt}</math> of photosynthesis</li> <li>Constructive adjustment:               <ul style="list-style-type: none"> <li>+ <math>A_{Opt}</math></li> </ul> </li> <li>Detractive adjustment:               <ul style="list-style-type: none"> <li>– <math>A_{Opt}</math></li> <li>– Respiratory capacity</li> <li>+ Isoprene emission</li> </ul> </li> <li>Change in morphology, e.g.               <ul style="list-style-type: none"> <li>– leaf mass per area<sup>c</sup></li> <li>+ Mesophyll conductance<sup>d</sup></li> </ul> </li> </ul>	<ul style="list-style-type: none"> <li>+ Heat tolerance</li> <li>– Low-temperature tolerance</li> <li>+ Isoprene emission<sup>h</sup></li> <li>Respiration homeostatic with pre-warming</li> </ul>		

(continued)

**Table 1** (continued)

	Biophysical response	Acclimation	Adaptation	Adaptation/Species turnover	Species turnover/speciation
<i>Sub-leaf/Organelle</i>	<ul style="list-style-type: none"> <li>+ Monoterpene synthesis</li> <li>+ Protein turnover</li> <li>+ Membrane fluidity</li> <li>+ Photorespiration</li> <li>- Electron transport rate</li> <li>- CO<sub>2</sub> solubility</li> </ul>	<ul style="list-style-type: none"> <li>+ Heat tolerance of photosynthetic machinery</li> <li>+ Thermostability of Rubisco activase or different isoforms being expressed<sup>e,f</sup></li> <li>+ Electron transport capacity</li> <li>+ Membrane rigidity Change in membrane composition (e.g., increased ratio of saturated to unsaturated fatty acids)<sup>g</sup></li> </ul>		If closed canopy forests are converted into savanna/desert ecosystems: increased abundance of C <sub>4</sub> and CAM species	
	<i>Instantaneous</i>	<i>Days-months</i>	<i>Years-decades</i>	<i>Decades-decennia</i>	<i>Decennia-Millenia</i>
<i>Temporal scale</i>					

Ecosystem and biome-level predictions for long timescales are speculative

<sup>a</sup>Zhang et al. (2014), <sup>b</sup>Fabry et al. (2015), <sup>c</sup>Cheesman and Winter (2013b), <sup>d</sup>Makino et al. (1994), <sup>e</sup>Salvucci and Crafts-Brander (2004), Wang et al. (2010), <sup>f</sup>Murakami et al. (2000), <sup>g</sup>Sharkey and Monson (2014)

## Direct Effects: CO<sub>2</sub> Fixation

At a given light level, photosynthesis is limited by one of three factors: (1) the capacity of ribulose 1,5-bisphosphate carboxylase/oxygenase (Rubisco) to carboxylate ribulose biphosphate (RuBP) (Rubisco-limited, or RuBP-carboxylation-limited photosynthesis); (2) the capacity to regenerate RuBP through the Calvin cycle and the thylakoid reactions (RuBP-regeneration-limited photosynthesis); or (3) the capacity for triose phosphate use (TPU) through starch and sucrose synthesis, where low TPU limits the regeneration of inorganic phosphate necessary for photophosphorylation (TPU-limited photosynthesis) (Harley and Sharkey 1991; von Caemmerer 2000). At light levels that saturate photosynthesis, RuBP-regeneration capacity generally reflects the maximum electron transport capacity ( $J_{\max}$ ), whereas the rate of RuBP-carboxylation-limited photosynthesis represents the maximum capacity for Rubisco carboxylase activity ( $V_{C\max}$ ). TPU limitation at current atmospheric CO<sub>2</sub> concentrations only occurs at low temperatures (Sage and Kubien 2007), and is expected to have minimal impact on photosynthesis in lowland tropical forest trees (but see Ellsworth et al. 2015 for potential TPU limitation driven by foliar phosphorus limitation). At current ambient CO<sub>2</sub> concentrations, RuBP-carboxylation limitation is common at intermediate temperatures, whereas RuBP-regeneration can become limiting at high temperatures (Sage and Kubien 2007). High temperature can also impede Rubisco functioning through its effect on Rubisco activase, the enzyme that promotes the dissociation of inhibitory sugar phosphates from the active site of Rubisco, thereby activating it (Portis 2003). Rubisco activase has lower heat tolerance than Rubisco itself, so high-temperature impairment of Rubisco activity may represent reduced activation of Rubisco, rather than reduced functioning of activated Rubisco (Salvucci and Crafts-Brandner 2004).

Rubisco can not only carboxylate RuBP, it can also oxygenate it, leading to photorespiration, a process that results in loss rather than gain of CO<sub>2</sub>. Photorespiration, although associated with reduced carbon gain, appears to play a beneficial role under high temperatures by maintaining electron flow and preventing photo-oxidation (Osmond and Björkman 1972; Kozaki and Takeba 1996), and by providing a substrate for the synthesis of isoprene (Jardine et al. 2014), a volatile organic compound believed to be associated with thermoprotection of photosynthesis (see “Volatile organic compounds and thermoprotection”). Increasing temperature promotes photorespiration in two ways. First, the solubility of CO<sub>2</sub> decreases more strongly with temperature than that of O<sub>2</sub>, so proportionally more O<sub>2</sub> can reach the active sites of Rubisco and stimulate RuBP oxygenation. Second, the relative specificity of Rubisco for CO<sub>2</sub> compared to O<sub>2</sub> decreases with increasing temperature, so at higher temperatures, higher CO<sub>2</sub> concentrations are needed to achieve a given RuBP-carboxylation rate (von Caemmerer and Quick 2000).

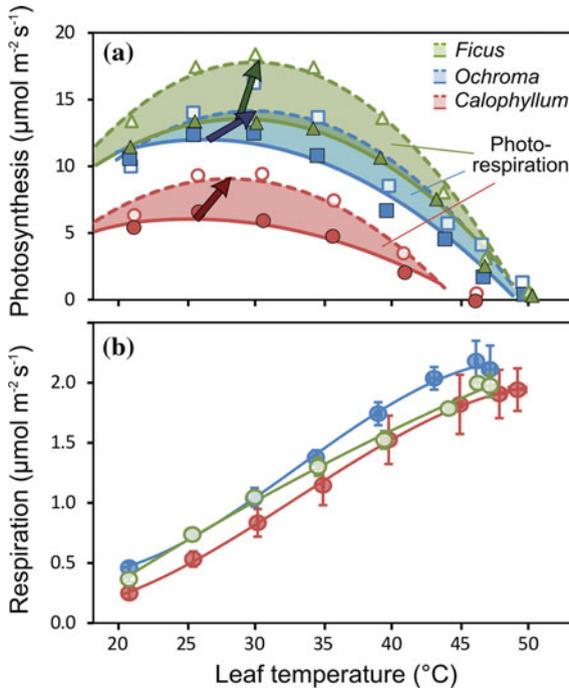
Under current ambient CO<sub>2</sub> concentrations, photosynthesis is controlled by TPU capacity at low temperature (< ~20 °C for the tropical species *Ipomoea batatas* (L.) Lam. (sweet potato; Sage and Kubien 2007). At intermediate temperatures RuBP

carboxylation becomes the rate-limiting step. With further warming ( $> \sim 32$  °C for *I. batatas*) increased photorespiration reduces photosynthetic efficiency, and the electron transport processes of RuBP regeneration become limiting. Ultimately, at temperatures exceeding the thermal tolerance of Rubisco activase, Rubisco activation may become limiting. Increasing CO<sub>2</sub> concentrations will reduce photorespiration, thus increasing the rates of both RuBP-regeneration-limited photosynthesis and RuBP-carboxylation-limited photosynthesis, without affecting TPU-limited photosynthesis. RuBP-carboxylation-limited photosynthesis is more strongly stimulated by CO<sub>2</sub> than RuBP-regeneration limited photosynthesis. Consequently, at higher CO<sub>2</sub>, there is a shift in what controls net photosynthesis at a given temperature, with control exerted by TPU extending to higher temperatures than at low CO<sub>2</sub>, and RuBP regeneration starting to limit photosynthesis at lower temperatures, resulting in a reduced or disappearing role of RuBP-carboxylation-limitation in constraining net photosynthesis (Sage and Kubien 2007). In other words, at elevated CO<sub>2</sub>, J<sub>max</sub> exerts stronger control over the temperature response of photosynthesis than V<sub>Cmax</sub>. The remainder of our discussion will focus primarily on temperature effects at current ambient CO<sub>2</sub> concentration ( $\sim 400$  ppm).

Figure 1a illustrates how temperature affects net CO<sub>2</sub> assimilation at 21 % versus 2 % O<sub>2</sub>—i.e., with and without photorespiration—in two early-successional tropical tree species, *Ochroma pyramidale* (Cav. ex Lam.) Urb. and *Ficus insipida* Willd., and in the late-successional species *Calophyllum longifolium* Willd. Because photorespiration is proportionally higher at higher temperatures (Slot et al. 2016), T<sub>Opt</sub> of net photosynthesis is higher at 2 % O<sub>2</sub>, similar to what has been observed under elevated CO<sub>2</sub> (e.g., Berry and Björkman 1980). *C. longifolium* has the lowest rates of photosynthesis regardless of temperature, consistent with its conservative growth strategy. *O. pyramidale* and *F. insipida*, in contrast, are fast growing species with higher T<sub>Opt</sub> and A<sub>Opt</sub> values. They germinate in forest gaps where irradiance levels as well as temperatures are higher than in the understory where *C. longifolium* typically regenerates.

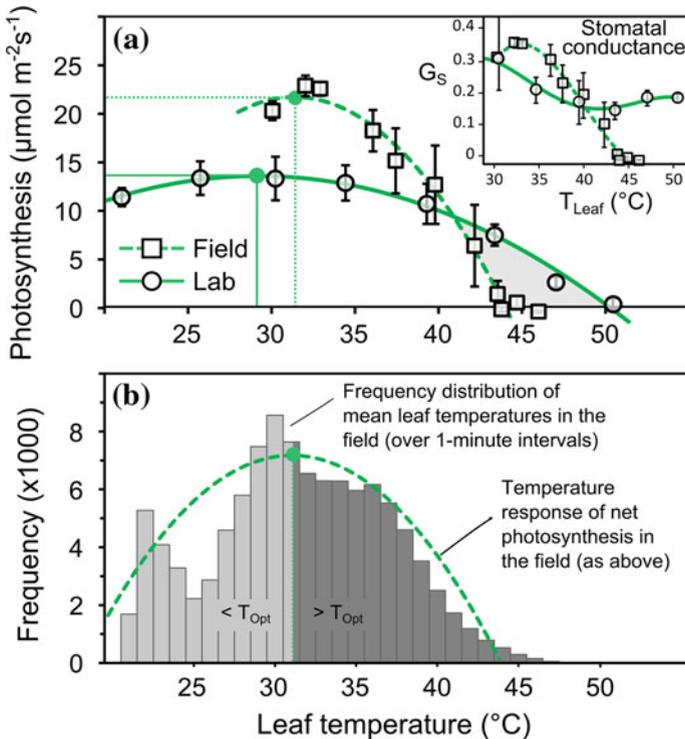
### Direct Effects: Thermodamage

One of the most temperature-sensitive components of plants is photosystem II (PSII), a protein complex located in the thylakoid membrane of the chloroplast. PSII efficiency can be readily determined using chlorophyll *a*-fluorescence techniques (e.g., Krause and Weis 1991). Thermodamage assessment from chlorophyll fluorescence measurements correlates well with classical analysis of post-heating necrosis (Krause et al. 2010, 2013). The critical temperature for irreparable leaf damage is about 52 or 53 °C, depending on whether thermal damage is determined in the dark or in the light (Krause et al. 2015). Such temperatures are only a few degrees above leaf temperatures that already occur occasionally in situ in sun-exposed leaves of tropical forest trees (Krause et al. 2010; Fig. 2). There is some variation across species in reported thermotolerance, but this may in part



**Fig. 1** Temperature response curves of net photosynthesis under photorespiratory (21 % O<sub>2</sub>, dashed lines, closed symbols), and non-photorespiratory conditions (2 % O<sub>2</sub>, solid lines, open symbols) **a**, and dark respiration **b** of recently fully expanded leaves of *Calophyllum longifolium* (circles), *Ficus insipida* (triangles), and *Ochroma pyramidale* (squares) seedlings grown outdoors under full natural radiation in Panama (mean annual temperature 27 °C). Arrows indicate increases in T<sub>Opt</sub> and A<sub>Opt</sub> with removal of photorespiration. Non-photorespiratory conditions were created by mixing air entering the cuvette with nitrogen gas at a 1:9.5 ratio. After passing the mixture through soda lime, CO<sub>2</sub> was added to generate a CO<sub>2</sub> concentration of 400 ppm. Measurements were made on attached leaves (n = 3–6) in a temperature-controlled Walz cuvette (Walz GmbH, Eifeltrich, Germany) attached to an LI-6252 infrared gas analyzer (Licor). Photosynthesis curves were fit as: net photosynthesis =  $b \times (T_{\text{Leaf}} - T_{\text{Min}}) \times (1 - e^{c \times (T_{\text{Leaf}} - T_{\text{Max}})})$ . T<sub>Min</sub> and T<sub>Max</sub> are the hypothetical low- and high-temperature CO<sub>2</sub> compensation points, and  $b$  and  $c$  are constants. All four variables were estimated using a non-linear solver function. Respiration fits are 3rd order polynomials. For clarity only means are shown in **a**; in **b** means ± 1 SEM are shown. Modified after Slot et al. (2016)

reflect differences in methodology used to assess thermotolerance. In a study comparing tropical and temperate rainforest tree species in Australia, the leaf temperature at which irreversible damage occurred was independent of growth temperature and biome of origin (Cunningham and Read 2006), suggesting minimal acclimation and phylogeographic predisposition to thermal damage.



**Fig. 2** Comparison of field and laboratory based measurements of net photosynthesis versus leaf temperature ( $T_{\text{Leaf}}$ ) of *Ficus insipida* **a**, and the frequency distribution of daytime  $T_{\text{Leaf}}$  for this species in the field **b** based on 1 month (Jan–Feb, dry season) of continuous monitoring of 6 leaves with thermocouple wires touching the abaxial surfaces. The *inset* in **a** shows the temperature response of stomatal conductance ( $G_s$ ). Lab measurements were made as described in Fig. 1. Field measurements were made with an LI-6400 (Licor) between 8 a.m. and noon on a typical dry season day. The leaves of the  $\sim 30$  m tall study tree were accessed using a canopy crane. The response *curve* of field-based measurements is overlaid on the frequency distribution of  $T_{\text{Leaf}}$  in **b** to illustrate the fact that  $T_{\text{Opt}}$  occurs at the most frequently occurring  $T_{\text{Leaf}}$ . *Error bars* represent 1 SEM ( $n = 4-6$ ). Modified after Slot et al. (2016)

**Indirect Effects: CO<sub>2</sub> Fixation**

Photosynthetic carbon uptake may also be affected by temperature via leaf-to-air vapor pressure deficit (VPD). Warm air can accommodate more water vapor than cool air, so at a given absolute moisture content of the air, VPD increases with leaf temperature, potentially causing stomatal closure. It is very difficult with standard gas-exchange equipment to keep VPD constant over a large range of leaf temperatures. One study that appears to have successfully controlled for VPD reported  $T_{\text{Opt}}$  values of 28–29 °C for two rainforest tree species in Costa Rica measured in situ (Vargas and Cordero 2013), values that are comparable to those reported in Fig. 1, in which VPD was not controlled. These observations also roughly correspond to

decreasing  $A_{\text{Net}}$  values at temperatures  $>30$  °C that Doughty and Goulden (2008) observed on three species measured in situ in the Brazilian Amazon, and with observations of Cheesman and Winter (2013a) of decreasing  $A_{\text{Net}}$  when  $T_{\text{Leaf}}$  was  $>27$ – $30$  °C in three tropical tree species grown in controlled-environment chambers in Panama. Whereas Vargas and Cordero (2013) maintained VPD  $< 2.0$  kPa at leaf temperatures above  $T_{\text{Opt}}$ , VPD in the experiment depicted in Fig. 1 increased from about 2.0 kPa at  $T_{\text{Opt}}$  to 8.0 kPa at the upper  $\text{CO}_2$  compensation point. Nonetheless, these two studies show similar decreases in net photosynthesis at temperatures above  $\sim 30$  °C. Insufficient data on VPD-controlled temperature responses of  $A_{\text{Net}}$  of tropical trees complicate the interpretation of the role of VPD-induced stomatal closure in determining  $T_{\text{Opt}}$  and photosynthetic decline above  $T_{\text{Opt}}$ .

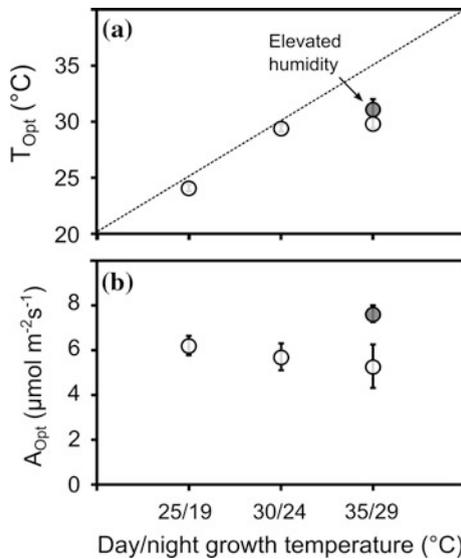
In upper-canopy tree leaves of tropical forest canopies, the combination of high temperature, high irradiance and high VPD can lead to pronounced midday depressions of stomatal conductance and  $\text{CO}_2$  uptake (e.g., Zotz et al. 1995; Cernusak et al. 2013; Goldstein et al. this volume, Santiago et al. this volume). Figure 2 shows changes in net  $\text{CO}_2$  uptake of *F. insipida* leaves measured in situ in the upper-canopy of a tropical forest in Panama as photosynthetic photon flux density (PPFD), temperature and VPD all increase from morning to noon on a sunny day. Compared to laboratory-measured seedling leaves (see Fig. 1 for methods),  $A_{\text{Opt}}$  of canopy leaves was much higher because these leaves were acclimated to higher PPFD and had higher leaf mass per area.  $T_{\text{Opt}}$  was also higher in the field, probably owing to generally higher leaf temperatures in situ. Whereas stomatal conductance and photosynthesis in the field decreased to zero when leaf temperatures reached 45 °C, in the laboratory net photosynthesis remained positive up to 50 °C and conductance—after a marked decrease between 30 and 45 °C—started to increase above 45 °C (Fig. 2). This re-opening of stomata at high temperatures in the laboratory was also observed for *O. pyramidale* and *C. longifolium* (Slot et al. 2016). Nonetheless, in exposed canopy leaves, high heat load caused by high solar radiation can become so stressful that photosynthetic carbon uptake may take place mainly during morning hours. During the course of the day depicted in Fig. 2a, 50 % of the time leaf temperatures exceeded  $T_{\text{Opt}}$  of *F. insipida* (Fig. 2b), stomata were partially closed, and  $A_{\text{Net}}$  was reduced. Depending on leaf orientation, the extent to which midday depression occurs may vary considerably, even between neighboring leaves, or within leaves if they are undulated (e.g., *Cecropia*; K. Winter, unpublished data). The variability in leaf-level irradiance and leaf temperature makes stomatal conductance particularly challenging to model and predict with high spatial and temporal resolution. Given the frequent occurrence of midday stomatal reductions in tropical trees on sunny days, further understanding of this phenomenon is warranted to better predict global carbon fluxes in a warming world.

## Acclimation

Thermal acclimation is a biochemical, physiological, or morphological adjustment by individual plants in response to a change in the temperature regime, which

results in an alteration in the short-term response to temperature (Smith and Dukes 2013). Thermal acclimation of photosynthesis commonly leads to a shift of the thermal optimum of photosynthesis towards the new growth temperature (Box 1). This shift results from a change in relative contribution of RuBP carboxylation and RuBP regeneration in controlling net photosynthesis. The optimum temperature of RuBP regeneration is higher than that of RuBP carboxylation (Kirschbaum and Farquhar 1984; Hikosaka et al. 2006). Warm-acclimated plants tend to have a lower  $J_{max}/V_{Cmax}$  ratio than cool-grown control plants (e.g., Bernacchi et al. 2003). When  $J_{max}/V_{Cmax}$  is low, RuBP regeneration—with its relatively high  $T_{Opt}$ — exerts greater control over net photosynthesis than when  $J_{max}/V_{Cmax}$  is high in cool-grown plants, and as a result  $T_{Opt}$  of net photosynthesis is higher in warm-acclimated plants (Hikosaka et al. 2006). If the  $J_{max}/V_{Cmax}$  ratio is not reduced by high temperature,  $T_{Opt}$  may still increase if the activation energy of  $V_{Cmax}$  increases more with growth temperature than that of  $J_{max}$  (Hikosaka et al. 2006).

Consistent with thermal acclimation,  $T_{Opt}$  of *C. longifolium* seedlings grown in controlled-environment chambers increased with growth temperature (Fig. 3), but

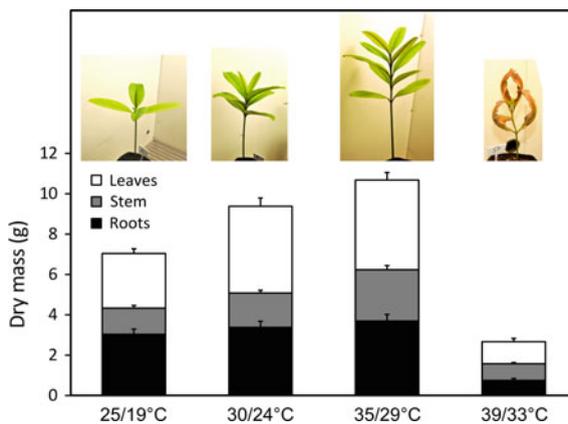


**Fig. 3** The optimal temperature of photosynthesis ( $T_{Opt}$ ) (a), and net photosynthesis at  $T_{Opt}$  ( $A_{Opt}$ ) (b) plotted against growth temperature of *Calopyllum longifolium* seedlings (mean  $\pm$  1 SEM). Seedlings were grown in individual pots (2.8 l) in commercial potting soil in growth cabinets (Environmental Growth Chambers, Chagrin Falls, OH, USA) at day/night temperatures of 25/19, 30/24, 35/29, 35/29 + elevated RH, and 39/33 °C. Plants from the 39/33 °C did not have enough live leaves for photosynthesis measurements (see Fig. 4). Net photosynthesis was measured with an LI-6400 (Licor) at 20, 25, 30, 35, and 40 °C on one leaf per seedling (n = 6/treatment) after equilibration for at least 45 min in growth cabinets set to the target temperature.  $T_{Opt}$  was calculated as described in the caption of Fig. 1. In one 35/29 °C cabinet relative humidity was increased to ~90 % (as opposed to ~45 %) by maintaining a pot of water near boiling point in the cabinet during the light period

there is a limit to this adjustment. Plants grown at 25/19 °C (day/night) and 30/24 °C had  $T_{Opt}$  values close to their daytime growth temperature, whereas plants grown at 35/29 °C had a  $T_{Opt}$  of 30 °C, suggesting that, at least for this late-successional species, 30 °C represents the maximum  $T_{Opt}$  at current  $CO_2$  concentrations. While  $T_{Opt}$  shifted towards the growth temperature,  $A_{Opt}$  decreased with increasing growth temperature. Way and Yamori (2014) call this *detractive* adjustment to warming, as opposed to *constructive* adjustment (see Box 1). The exception to this pattern was the 35/29 °C treatment associated with elevated relative humidity ( $\sim 90\%$  as opposed to  $\sim 45\%$ ), for which both  $T_{Opt}$  and  $A_{Opt}$  were highest among all treatments. This may indicate that part of the decrease in  $A_{Opt}$  relative to growth temperature resulted from temperature effects on VPD, with higher stomatal conductance being maintained when plants were grown at elevated relative humidity.

Kositsup et al. (2009) grew *Hevea brasiliensis* Müll. Arg. seedlings at 18 and 28 °C and also found that  $T_{Opt}$  adjusted to growth temperature. They further observed that  $A_{Net}$ ,  $V_{Cmax}$  and  $J_{max}$  all increased with increased growth temperature, suggesting constructive acclimation. However, 18 °C is probably sub-optimal for lowland tropical species such as *H. brasiliensis*, and the observed adjustments may indicate that 28 °C is closer to the optimum temperature for this species, rather than indicate strong constructive thermal acclimation. Doughty (2011) warmed leaves of trees and lianas (woody vines) in the Brazilian Amazon and found no acclimation of  $A_{Net}$ . The decrease in photosynthesis with warming was assigned to leaf damage caused by occasionally very high leaf temperatures of experimentally warmed leaves during sunny spells. Similarly, *C. longifolium* plants grown in controlled-environment chambers at 39/33 °C (day/night) showed severe leaf damage and photosynthesis could not be measured (Fig. 4).

Dusenge et al. (2015) reported that  $V_{Cmax}$  was similar between leaves of trees from two tropical rainforest sites in Rwanda differing in temperature regime,



**Fig. 4** The effect of growth temperature on total biomass (mean  $\pm$  1 SEM of 6 seedlings per treatment) and biomass allocation in *Calophyllum longifolium* seedlings, with photographs of representative plants of each treatment at final harvest

whereas  $J_{\max}$  was significantly lower at the warmer site, resulting in a reduced  $J_{\max}/V_{C\max}$ , in line with observations on temperate species. The physiological plasticity notwithstanding,  $A_{\text{Net}}$  was reduced at the warmer site compared to the cool site, suggesting detractive adjustment, similar to observations on *C. longifolium* (Fig. 3). We still know little about the thermal acclimation potential of  $V_{C\max}$  and  $J_{\max}$  of lowland tropical species, and to date no study has compared the acclimation potential of  $V_{C\max}$  and  $J_{\max}$  of tropical and temperate tree species. Such experimental studies are needed to better predict the effect of long-term warming on the photosynthetic properties of tropical forest vegetation.

## Adaptation

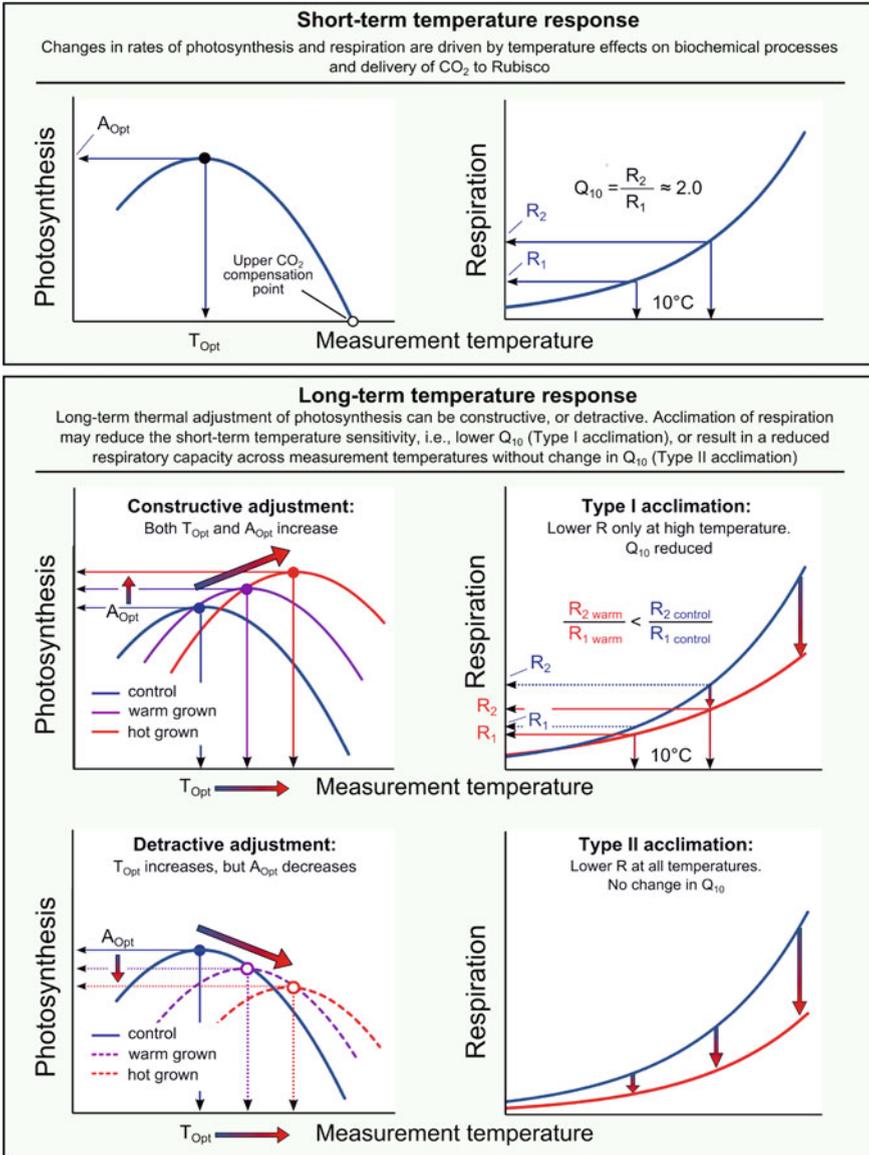
There is at least one comparative study that points to fundamental differences in the physiological properties of species adapted to temperate versus tropical conditions. Cunningham and Read (2002) reported higher  $T_{\text{Opt}}$  values for tropical than temperate species at a given growth temperature. Furthermore, the temperature response curve for temperate species was wider, i.e., temperate species had photosynthesis rates of  $>80\%$  of  $A_{\text{Opt}}$  over a larger temperature range. These results are consistent with the absence of strong selection for a broad temperature range of photosynthesis for tropical species. We found that  $T_{\text{Opt}}$  of tropical species is lower than growth temperature when growth temperatures exceed  $\sim 30\text{ }^{\circ}\text{C}$  (Fig. 3). The narrower curves of tropical species and the apparent limit to increasing  $A_{\text{Opt}}$ , despite a small increase in  $T_{\text{Opt}}$ , suggest that significant warming in the tropics may cause a decline in photosynthetic carbon uptake.

## Dark Respiration

### Temperature Response

About 30 % of the carbon fixed by tropical forests through photosynthesis is released back into the atmosphere by foliar respiration (Chambers et al. 2004; Malhi 2012). Respiration is vital for plant growth and survival, supporting biosynthesis, cellular maintenance and repair, but from a carbon-balance perspective respiration represents a loss.

In the short term, dark respiration increases with temperature, peaks with a  $T_{\text{Opt}} > 50\text{ }^{\circ}\text{C}$ , and then steeply declines. The initial increase is often assumed to be exponential, and can be expressed as a  $Q_{10}$  value—the proportional increase in respiration with  $10\text{ }^{\circ}\text{C}$  warming (Box 1). The general assumption is that respiration rates double for every  $10\text{ }^{\circ}\text{C}$  warming, resulting in a  $Q_{10}$  of 2.0, although in reality a wide range of  $Q_{10}$  values—from  $\sim 1.4$  to 4.2—has been reported (Atkin et al. 2005, and references therein). Leaf respiration of tropical forest species is at least as sensitive to short-term temperature increase as that of temperate species, with  $Q_{10}$



**Box 1.** Photosynthesis and respiration are highly sensitive to temperature, but the effects depend on the timescale of exposure. Long term warming (e.g., weeks to months) may modify the short-term response through acclimation, i.e., the biochemical, or structural adjustment by individual plants in response to a new temperature regime

values ranging from 1.5 to 4.1 (Meir et al. 2001; Cavaleri et al. 2008; Slot et al. 2013, 2014b), and averaging above 2.0 (Slot et al. 2013, 2014b). Figure 1b shows respiration-temperature response curves for leaves of *C. longifolium*, *F. insipida*, and *O. pyramidale*. Notably, the response-curves ( $n = 3-6$ ) are not exponential. Over a relatively narrow temperature range it is difficult to distinguish different shapes of temperature response curves, but the examples in Fig. 1b cover a range of  $\sim 25$  °C, and an exponential curve is clearly not an appropriate approximation. We do not know whether the data shown in Fig. 1b reflect a uniquely tropical phenomenon, but a recent study with *Oryza sativa* L. (rice) reported similarly linear temperature responses of respiration over a  $\sim 14$  °C range (Peraudeau et al. 2015). In contrast, measurements over a broad temperature range on the temperate *Eucalyptus pauciflora* suggest a good exponential fit up to critical temperatures around 50 °C (O’Sullivan et al. 2013), whereas Hüve et al. (2012) showed considerable variation in the shape of the response curve over a  $\sim 25$  °C range across three temperate herbs. Modeling temperature responses using an exponential  $Q_{10}$  or Arrhenius-type fit when a linear fit better describes the actual response could lead to overestimation of respiration rates at high temperatures. Future research will need to assess how widely distributed the non-exponential temperature responses of dark respiration are among tropical species.

## Acclimation

Nighttime temperatures increase more rapidly than daytime temperatures (e.g., Easterling et al. 1997), potentially leading to increased respiration costs in tropical forest systems. However, acclimation of respiration might mitigate this. Respiration at a given temperature is generally decreased in warm-acclimated plants compared to control plants, either through down-regulation of respiratory capacity at all temperatures, or because the short-term temperature sensitivity in warm-acclimated plants is reduced, leading to lower rates at intermediate to high temperatures (Atkin and Tjoelker 2003; see Box 1). Recently, we showed that upper-canopy leaves of tropical canopy trees and lianas acclimate to nighttime warming (Slot et al. 2014a). Acclimation has also been shown in seedlings of tropical tree species grown under different temperature regimes in controlled-environment chambers and open-top chambers (Cheesman and Winter 2013b). In fact, when accounting for the degree and duration of warming there is no indication that tropical species differ from species from other biomes in their capacity for thermal acclimation of respiration (Slot and Kitajima 2015).

At the whole-plant level, respiratory acclimation can be affected by interactions with mycorrhizal symbionts (Atkin et al. 2009). The only study to date that has addressed these interactions for tropical tree species indicated a strong species-specific effect of mycorrhizal colonization of roots on acclimation of whole-plant respiration to nighttime warming (Fahey et al. 2016). The host species-specific nature of the effects of mycorrhizae may complicate generalizations of acclimation trends in vegetation models, and calls for more research into the mechanisms behind these interactions.

## Adaptation

Respiration rates at ambient temperature vary considerably across biomes, with rates increasing twofold from the Arctic to the tropics (Atkin et al. 2015). This doubling over a  $\sim 20$  °C range in growing season temperature represents a considerable deviation from the much steeper instantaneous temperature response. Respiration standardized to 25 °C is threefold lower in the tropics than in the Arctic (Atkin et al. 2015), suggesting significant downregulation of respiratory capacity in warm-adapted species. These global patterns may reflect adaptation to different climates, but model simulations by Vanderwel et al. (2015) show that accounting for acclimation to local temperature regimes produces very similar patterns in simulated respiration across global temperature gradients. Given that the capacity for acclimation is common across the globe (Slot and Kitajima 2015) and sufficient to explain global patterns, it appears that there may not have been further selective pressure leading to adaptive temperature responses of respiration.

## *Volatile Organic Compounds and Thermoprotection*

Volatile organic compounds (VOC) can contribute significantly to the carbon emissions from forests. The most common VOCs are isoprenes and monoterpenes, and the emission of both increases with short-term warming in the tropics (Keller and Lerdau 1999; Bracho-Nuñez et al. 2013). Isoprene emission is hypothesized to protect photosynthesis from short, high-temperature episodes (Sharkey and Yeh 2001; Sasaki et al. 2007) by helping stabilize thylakoid membranes (Velikova et al. 2011), quenching reactive oxygen species (Velikova et al. 2012; Jardine et al. 2013, 2014), and/or increasing heat dissipation (Sasaki et al. 2007; Pollastri et al. 2014). In the short term (minutes to hours), isoprene emission increases exponentially with temperature, both in temperate (Sharkey and Monson 2014) and tropical trees (Keller and Lerdau 1999), but above 38–40 °C emissions may decrease (Tingey et al. 1979; Alves et al. 2014; Jardine et al. 2013, 2014).  $T_{\text{Opt}}$  of isoprene emission is  $\sim 10$  °C higher than  $T_{\text{Opt}}$  of net photosynthesis. Consequently, at high temperatures isoprene synthesis requires carbon sources other than recent photosynthates. Photorespiration may provide such an alternative source at temperatures exceeding  $T_{\text{Opt}}$  of photosynthesis (Jardine et al. 2014). Controlled-environment studies with temperate species have shown that plants acclimated to higher growth temperature have a higher capacity for isoprene release than plants grown at moderate temperatures (Tingey et al. 1979; Monson et al. 1992), but there is currently no evidence to suggest that warm-adapted tropical species have higher emission rates than temperate species.

Despite its potential importance in thermoprotection, isoprene emission does not occur in all plants. It only occurs in perennial plants with  $C_3$  photosynthesis (Loreto and Fineschi 2015), but without a clear phylogenetic distribution, due in part to frequent loss and secondary acquisition of the trait (Monson et al. 2013). In the tropics isoprene emission is equally common in deciduous and evergreen plants,

whereas emission is comparatively rare in temperate evergreens (Loreto and Fineschi 2015). Tambunan et al. (2006) studied isoprene emission in 42 tropical tree species and reported high rates in 4 species, very low rates ( $<10 \mu\text{g g}^{-1} \text{h}^{-1}$ ) in 28 species, and non-detectable rates in 10 species. Of the 51 tropical tree and lianas studied by Keller and Lerdau (1999), only 15 emitted isoprene at detectable rates. Even for the latter species, the total contribution of emission of isoprene to the carbon balance was relatively small, with an overall mean rate of  $24 \text{ nmol m}^{-2} \text{ s}^{-1}$  ( $\sim 65 \mu\text{g g}^{-1} \text{h}^{-1}$  at mean leaf mass per area of  $90 \text{ g m}^{-2}$ ) at  $30^\circ\text{C}$  and PPFD of  $1000 \mu\text{mol m}^{-2} \text{ s}^{-1}$ , which is  $\sim 500$  times lower than the mean photosynthesis rate. It is likely that long-term warming will increase rates of isoprene emission from leaves of isoprene-emitting species (Sharkey and Monson 2014), but given the thermoprotective role these emissions appear to play, the loss of isoprene carbon is probably more than compensated for by the maintenance or recovery of photosynthetic functioning during episodic heat.

## Whole-Plant Temperature Effects

### *Biomass Accumulation*

Two meta-analyses suggest that plant growth decreases with warming when starting temperatures are already high (Way and Oren 2010; Lin et al. 2010); in both studies tropical species were underrepresented. Figure 4 shows the effect of combined day and nighttime warming on growth of *C. longifolium*. Growth was optimal at  $35/29^\circ\text{C}$  (day/night), i.e., at temperatures well above those in its natural habitat in the forest understory. Several studies show that nighttime warming stimulates growth in seedlings of tropical tree species (Esmail and Oelbermann 2011; Cheesman and Winter 2013a, b). These seedling responses contrast with results obtained for tropical canopy trees, which suggest a negative correlation between nighttime temperature and tree growth (Clark et al. 2003, 2010, 2013; Vlam et al. 2014; Anderegg et al. 2015a). A recent study on *Eucalyptus* species across a temperature gradient in Australia reported that, compared to small trees, big trees show a disproportionately strong decline in growth with increasing mean annual temperature (Prior and Bowman 2014). Perhaps ontogenetic differences in temperature sensitivity govern the contrasting temperature effects on tropical canopy trees and seedlings. Leaf area ratio (leaf area divided by total plant mass) decreases with ontogeny, so canopy trees have proportionally less photosynthetic material with which to support the maintenance of non-photosynthetic tissue than seedlings. If the capacity for thermal acclimation of root and stem respiration is smaller than that of leaf respiration, then warming would increase the respiration load in canopy trees more strongly than in seedlings. There is no indication that acclimation of root respiration is lower than that of leaf respiration in temperate species (Loveys et al. 2003), but little is known about thermal acclimation of root and stem respiration in the tropics beyond the observation by Fahey et al (2016) that acclimation of whole-plant- and root respiration varied greatly among species.

In in situ studies of growth-temperature relationships—using latitudinal or elevational temperature gradients, or inter-annual trends in temperature—temperature is one of multiple factors that change, and patterns of biological response cannot be assigned to temperature differences alone. Controlled-environment chambers, on the other hand, enable the isolated study of temperature effects. However, in controlled-environment studies with potted seedlings, soil temperature may track changes in air temperature more closely than in the field, and soil warming itself may increase growth (Königer and Winter 1993; Holtum and Winter 2014). Forest soils are thermally very stable, so soil processes may explain at least part of the observed contrasting responses of seedlings and canopy trees to nighttime warming. Soil warming may increase nutrient mineralization rates (Jarvis and Linder 2000), thereby stimulating growth. However, we found that nighttime warming stimulated growth of *O. pyramidale* seedlings equally in fertilized and non-fertilized soils (Slot and Winter unpublished data). Altered source-sink relationships (Pilkington et al. 2015) may provide an alternative, though not yet tested explanation for the observed responses.

## ***Mortality***

A quantitative trait locus (QTL) for thermotolerance has been identified for the comparatively heat-tolerant *Oryza glaberrima* Steud. (African rice), the overexpression of which in *O. sativa* L., *Arabidopsis thaliana* L., and *Festuca arundinacea* Schreb., significantly reduced their mortality following a 12 day 38/35 °C treatment (Li et al. 2015). The QTL is associated with degradation of cytotoxic denatured proteins that accumulate during heat exposure, suggesting an important role for the proteins encoded by the QTL in mitigating warming-induced cell death and plant mortality. Several other factors may play a role in warming-induced mortality. For example, increased rates of respiration and decreased rates of photosynthesis could lead to carbon starvation. Atmospheric drought caused by temperature-induced increase in VPD may also kill plants. It is hard to disentangle drought and temperature effects, as soil moisture availability may prevent warming-induced carbon starvation by enabling transpirational leaf cooling and maintenance of a positive carbon balance during heat events (Bauweraerts et al. 2014), and carbon reserves in the form of soluble carbohydrates may prevent drought-induced hydraulic failure (O'Brien et al. 2014). Drought is a common agent of mortality in the tropics (e.g., Condit et al. 1995; Allen et al. 2010), subject of observational (e.g., Condit et al. 1995; Phillips et al. 2010) and experimental studies (e.g., Slot and Poorter 2007; Nepstad et al. 2007), but atmospheric drought, such as occurs during warming, is much less studied (Breshears et al. 2013).

In the *C. longifolium* study depicted in Fig. 4, daytime temperature of 39 °C and relative humidity of ~29 % was lethal for most seedlings. We do not know

whether the well-watered seedlings died from cytotoxicity, carbon starvation, or hydraulic failure, but the fact that  $T_{Opt}$  and  $A_{Opt}$  at 35/29 °C were higher under elevated relative humidity (Fig. 3), shows that atmospheric drought can negatively impact the maintenance of a positive carbon balance, and potentially survival. Tropical sapling mesocosms grown outdoors in Panama, with daytime warming of ~5 °C did not negatively impact biomass accumulation and photosynthetic traits as long as plants had access to soil water (Slot and Winter unpublished data), further highlighting the interconnected nature of heat and drought as agents of plant mortality.

Elevated CO<sub>2</sub> improves both the carbon balance and water use efficiency of plants, but a recent greenhouse study on *Eucalyptus radiata* A.Cunn. ex DC suggests that negative effects of elevated temperature on drought-induced mortality are not alleviated by elevated CO<sub>2</sub>, despite higher leaf-level water use efficiency in the high CO<sub>2</sub> treatment (Duan et al. 2014). Thus, drought-induced mortality may occur in parts of the tropics where warming is accompanied by decreased precipitation (Olivares et al. 2015). Furthermore, abiotic climatic changes are likely to interact with biotic sources of mortality such as insect outbreaks (Anderegg et al. 2015b). For example, warming can affect phytochemistry and the feeding efficiency of invertebrate herbivores (Jamieson et al. 2015), potentially leading to increased leaf loss and plant mortality.

## **Reproduction**

There have not been any detailed experimental studies on how rising temperatures affect reproduction of tropical vegetation, although the reproductive phase is one of the most temperature-sensitive parts of the lifecycle of plants. Pollen production, pollen viability and pollen tube growth all decrease with warming in a range of crop species (Sage et al. 2015, and references therein). There is considerable variation in temperature sensitivity across genotypes and species, but no clear relationship between ambient growth environment and thermal sensitivity (Prasad et al. 2006). Pollen viability of the tropical tree *Mangifera indica* L. (mango) decreases above 33 °C (Issarakraisila and Considine 1994), which is not an unusually high temperature in the tropics. Many tropical species flower for only short periods and risk reproductive failure if flowering coincides with anomalously high temperatures. Surprisingly, flower production correlated positively with temperature both seasonally and interannually in tropical forests in Panama and Puerto Rico (Pau et al. 2013). Increased flower production could compensate for decreased pollen viability but detailed study is required to better understand temperature effects on tropical tree reproduction and to disentangle direct warming effects on plant fertility, and warming effects on reproductive allocation.

Temperature may also affect reproduction through its effect on pollinators. For example, fig-pollinating wasps—which develop inside developing figs—die at

temperatures only a few degrees above current ambient temperatures (Patiño et al. 1994). Transpirational cooling helps maintain fruit temperatures in non-lethal ranges, but further warming could have major negative consequences for fig reproduction.

Exposure to high temperature is necessary to break physical dormancy of seeds of many species. This is especially common for species that form seed banks, as the high temperature is associated with high light conditions favorable for seedling establishment of these early-successional species. More intense heatwaves (Meehl and Tebaldi 2004) could potentially trigger germination under conditions not favorable for seedling establishment, leading to mortality. The potential effects of climate change on the seed ecology of tropical forest trees is not well understood (Walck et al. 2011).

## Stand-, Ecosystem-, and Biome-Level Temperature Effects

An important aspect of tropical forests is their very high species diversity. All trees ultimately use—and often directly compete for—the same resources, and species coexistence is at least in part maintained by niche specialization along resource gradients (e.g., Kitajima and Poorter 2008; Condit et al. 2013). This means that climate warming may shift the competitive balance within a forest community if rising temperature differentially affects mortality and reproduction across species. For example, warming may increase soil mineralization rates in the tropics (Salinas et al. 2011). Tropical tree species vary widely in their phosphorus affinity (Condit et al. 2013), and temperature-induced changes in nutrient availability potentially contribute to shifts in species composition. This may result in reduced diversity, or a change in the dominant species and functional types associated with a particular nutrient regime. Climate-induced changes in species composition will affect forest growth (Coomes et al. 2014). Systematic changes in growth rates of trees may also cause turnover rates in tropical forests to increase (Phillips and Gentry 1994). Nevertheless, changes in tree growth rates and turnover do not necessarily change ecosystem carbon storage if the size structure of trees within the ecosystem is maintained (Körner 2009).

Tropical ecosystems have lower ratios of NPP over gross primary productivity (GPP) than most higher latitude ecosystems (Zhang et al. 2009, 2014). Furthermore, rising temperatures decrease the NPP/GPP ecosystem carbon use efficiency (Zhang et al. 2014) suggesting that, despite observed thermal acclimation capacity at the leaf level, warming causes ecosystem-level respiration cost to increase. Soil respiration is a major component of ecosystem respiration, and soil respiration rates in tropical forests are higher than in any other ecosystem in the world (Raich and Schlesinger 1992). The capacity of tropical soil respiration to acclimate to higher temperatures is unknown.

## Concluding Remarks

Because most research on temperature effects on the ecophysiology of plants has been done on non-tropical species, much is still to be discovered about how tropical trees will respond to climate change. We have a fairly sound understanding of general principles of temperature responses at the organelle and leaf level (Table 1), but we do not know whether tropical trees differ systematically from non-tropical species in how warming affects leaf and whole-plant performance. For example, more research is needed to establish the generality of detractive acclimation of photosynthesis in tropical species, as observed in *C. longifolium* (Fig. 3). We have shown that the commonly assumed exponential increase of respiration with temperature may not apply to tropical leaves over wide temperature ranges, a finding that requires further confirmation. At this point, we can only speculate on the whole tree-level effects of warming, apart from noting that warming is associated with atmospheric drought and may affect reproduction. At the stand level, warming may exacerbate midday reductions in stomatal conductance. A better mechanistic understanding of the causes and consequences of midday stomatal closure will improve our ability to translate understanding based on individual leaf responses to stand-level photosynthesis simulations.

In the decades to come the tropics will experience further increases in atmospheric CO<sub>2</sub> concentrations and warming, and significant alterations in the timing and amount of rainfall (Diffenbaugh and Scherer 2011; Mora et al. 2013). The vulnerability of tropical forests to climate change is under much debate (e.g., Clark 2004; Lloyd and Farquhar 2008, Lewis et al. 2009; Booth et al. 2012; Randerson 2013). Because of the large contribution of tropical forests to the global carbon cycle, a better understanding of how these forests respond to climate change drivers will improve our ability to predict future climate and biogeochemical cycling at the global scale (Booth et al. 2012; Huntingford et al. 2013; Piao et al. 2013). Information on thermal acclimation of photosynthetic parameters  $V_{C_{max}}$  and  $J_{Max}$  will be crucial to improve predictive models De Kauwe et al. (2016). We can learn a lot about plant-temperature interactions from elevation gradients in the tropics (e.g., Malhi et al. 2010), but these gradients do not reveal how plants at the high-temperature end of the gradient will respond to further warming, and that is precisely the key in determining the fate of tropical lowland forests in a warming world. As global warming generates novel climate regimes in tropical latitudes, maintenance of current forest structure requires processes of thermal acclimation to occur in conjunction with adjustments to changing precipitation regimes (Mora et al. 2013)—and possibly changes in cloud cover—under gradually rising ambient CO<sub>2</sub> concentrations and increasing atmospheric deposition of nutrients (Hietz et al. 2011). Clearly, in situ warming experiments of entire forest segments are needed to better assess the effects of rising temperatures on tropical vegetation (Cavaleri et al. 2015).

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